

Cell Measurement Protocol

1. Transform *Escherichia coli* TOP10 with these following plasmids:

Positive control

Negative control

Device 1: J23101+I13504

Device 2: J23106+I13504

Device 3: J23117+I13504

2. Pick 2 colonies from each of plate and inoculate it on 5-10 mL LB medium +

Chloramphenicol.

Grow the cells overnight (16-18 hours) at 37°C and 220 rpm

3. Cell growth, sampling, and assay

- Set your instrument to read OD600 (as OD calibration setting)
- Measure OD600 of the overnight cultures
- Record data in the notebook
- Import data into Excel (normalisation tab) Sheet_1 provided
- Dilute the cultures to a target OD600 of 0.02 in 10 ml LB medium + Chloramphenicol in 50 mL falcon tube

- Take 100 μL (1% of total volume) samples of the cultures in the 96-well plates, Use Thermo VARIOSKAN FLASH to shake the 96-well plates, measure the samples (OD and FITC) at 0, 1, 2, 3, 4, 5, and 6 hours of incubation
- Record data in the notebook
- Import data into Excel (cell measurement tab) Sheet_1 provided